

Synthesis, Biological Properties, and Molecular Modeling Investigation of the First Potent, Selective, and Water-Soluble Human A₃ Adenosine Receptor Antagonist

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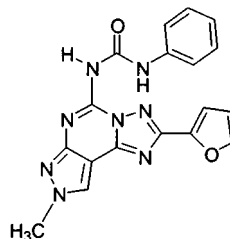
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Abstract: A new, highly potent, selective, and water-soluble antagonist of the hA₃ adenosine receptor was synthesized and tested in binding and functional assays. Compound **4** (5-[[4-(pyridyl)amino]carbonylamino-8-methyl-2-(2-furyl)-pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine hydrochloride) displayed high water solubility (15 mM) and the highest affinity ($K_i = 0.01$ nM) and selectivity for the hA₃ versus A₁, A_{2A}, and A_{2B} receptors (>10000-fold) ever reported. A Schild analysis of the antagonism by **4** of agonist-induced inhibition of cAMP production in CHO cells expressing the hA₃ receptor indicated a K_B value of 0.20 nM.

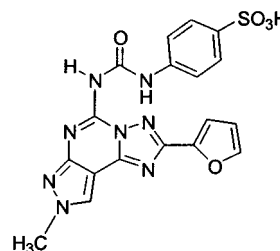
Introduction. It is well-known that adenosine regulates many physiological processes through the interaction with four known receptor subtypes classified as A₁, A_{2A}, A_{2B}, and A₃.^{1,2} In particular, the A₃ adenosine receptor subtype, which is distributed in different organs (lung, liver, kidney, heart, and, with a lower density, the brain),³ exerts its action through the modulation of two second messenger systems: inhibition of adenylate cyclase⁴ and stimulation of phospholipases C⁵ and D.⁶ The potential therapeutic applications of activating or antagonizing this receptor subtype have been investigated in recent years. In particular, antagonists for the A₃ receptor promise to be useful for the treatment of inflammation⁷ and in the regulation of cell growth.^{8,9} Consequently, much effort has been directed toward searching for potent and selective human A₃ adenosine antagonists.¹⁰ Recently, Baraldi and co-

Chart 1. Structures, Biological Characterization, and Water Affinities of Reference Compounds



1

hA₁ K_i = 594 nM; hA_{2A} K_i = 381 nM
hA_{2B} K_i = 222 nM; hA₃ K_i = 0.16 nM
hA₁/hA₃ = 3,713; hA_{2A}/hA₃ = 2,381
hA_{2B}/hA₃ = 1,388
R_m = 4.06 ± 0.05



2

hA₁ K_i = >10,000 nM; hA_{2A} K_i = 594 nM
hA_{2B} K_i = >10,000 nM; hA₃ K_i = 25 nM
hA₁/hA₃ = > 400; hA_{2A}/hA₃ = 24
hA_{2B}/hA₃ = > 400
R_m = 1.66 ± 0.18

workers reported a large series of pyrazolotriazolopyrimidines, bearing substituted phenylcarbonyl residues at the amino group at the 5-position, as highly potent and selective antagonists of the human A₃ adenosine receptor.^{11–13} In particular, **1** (5-[[4-(phenylamino)carbonylamino-8-methyl-2-(2-furyl)-pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine hydrochloride) (Chart 1) showed the most favorable binding affinity and selectivity for the human A₃ adenosine receptor ever reported.¹³

Unfortunately, a major problem within this class of compounds is the typical low water solubility, which has limited their use as pharmacological and diagnostic tools. The hydrophobicity of compound **1** is indicated by its high R_m value of 4.06. Previously an attempt for obtaining a water-soluble derivative has been made through the introduction of a sulfonic acid group at the para position of the phenyl ring (Chart 1) affording compound **2** (4-[3-(2-furyl-2-yl-8-methyl-8H-pyrazolo[4,3-*e*][1,2,4]triazolo[1,5-*c*]pyrimidin-5-yl)ureido]benzenesulfonic acid). As expected, a completely water-soluble derivative ($R_m = 1.66$, water solubility greater than 20 mM) was obtained; however, a significant loss of affinity (156-fold) and selectivity was observed.¹³ A hypothesis for the dramatic loss of affinity was provided through molecular modeling studies, which indicated that steric control seemed to be taking place around the para position of the phenyl ring in the putative A₃ receptor binding site.¹³ Taking into account these observations and with the aim of obtaining derivatives with high affinity, selectivity, and water solubility, we synthesized

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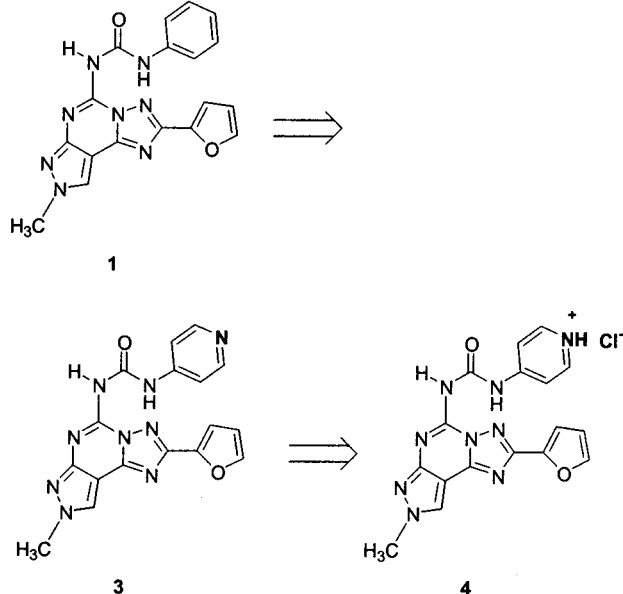
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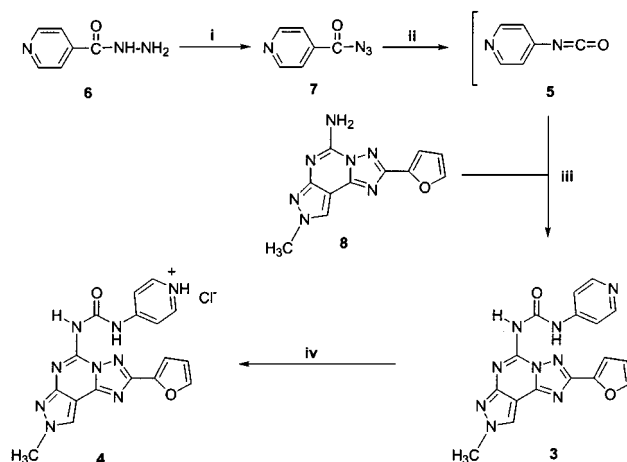
Chart 2. Rational Design of Water-Soluble hA₃ Adenosine Receptor Antagonist

a new derivative (Chart 2) by bioisosteric replacement of the phenyl ring with the 4-pyridyl moiety (**3**), thus providing higher water solubility while avoiding the steric hindrance of a substituent at the para position, which seemed to be responsible for the reduction of hA₃ adenosine receptor affinity.

In fact, the introduction of a basic nitrogen was intended to further improve water solubility when protonated, i.e., in the form of the corresponding HCl salt (**4**).

Results and Discussion. The synthesis of the desired compound has been performed following the general strategy depicted in Scheme 1. The major problem in the synthesis of derivative **3** involved the preparation of a 4-pyridyl isocyanate (**5**), which could not be synthesized by the usual method, i.e., reaction of the corresponding amine with phosgene or a phosgene equivalent, because of the reactivity and instability of pyridyl isocyanates.¹⁴

For this reason, **5** was prepared as reported in the literature starting from the commercially available isonicotinoyl hydrazide (**6**), which after reaction with sodium nitrite under acid conditions afforded the corresponding acyl azide **7**.¹⁵ The latter was in turn converted into **5** upon Curtius rearrangement induced by heating **7** at reflux in dry benzene for 2 h.¹⁶ The crude isocyanate was refluxed overnight in dry THF with the precursor containing the tricyclic system bearing a methyl group at the N8 position (**8**). The desired product **3** was purified using flash chromatography and eluted

Scheme 1^a

^a (i) NaNO₂, aqueous HCl, 0 °C, 1 h; (ii) benzene, reflux, 2 h; (iii) THF reflux overnight; (iv) HCl/MeOH, 0 °C, 30 min.

with a MeOH/EtOAc gradient of 0–30%. The corresponding hydrochloride **4** was obtained by treatment of **3** for 30 min at 0 °C with methanol saturated with HCl gas. The hydrophobicity of the newly synthesized substances was measured in reverse-phase TLC experiments and reported as *R_m* values¹⁷ (*R_m* = log(1/*R_f* – 1)) as shown in Table 1. As expected, both derivatives showed increased hydrophilicity with respect to reference **1**, but most importantly, the hydrochloride salt **4** freely dissolved in water to a maximum concentration of 15 mM. Table 1 also compares the receptor binding affinities of **3** and **4** determined at human A₁,¹⁸ A_{2A},¹⁹ A_{2B},²⁰ and A₃²⁰ receptors expressed in CHO (A₁, A_{2A}, A₃) and HEK-293 (A_{2B}) cells.

Surprisingly, both substances showed very high affinity at the human A₃ adenosine receptor subtype, with *K_i* values in the picomolar range (10–40 pM) and with high levels of selectivity. In particular, the hydrochloride salt **4** showed increased affinity and selectivity with respect to the reference compound bearing the phenyl-carbamoyl moiety **1**. The *K_i* value of **4** at the hA₃ receptor was 0.01 nM, thus indicating high selectivity versus other subtypes: hA₁/hA₃ = 35 000, hA_{2A}/hA₃ = 10 000, hA_{2B}/hA₃ = 25 000. These values were more favorable than the selectivity ratios for compound **1**, for which the *K_i* value at the hA₃ was reported to be 0.16 nM: hA₁/hA₃ = 3700, hA_{2A}/hA₃ = 2400, hA_{2B}/hA₃ = 1400. Moreover, this class of compounds, as previously demonstrated, proved to be inactive in a rat model with *K_i* values at the rA₃ typically greater than 1 μM.¹¹ **4** at 1 μM displaced only 35% of the specific binding of [¹²⁵I]-AB-MECA at A₃ receptors in membranes of rat basophilic RBL-2H3 cells.^{21b} Concerning the affinity differences observed between the salt and neutral species, we

Table 1. Binding Affinity at hA₁, hA_{2A}, hA_{2B}, and hA₃ Adenosine Receptors and Water Solubility of Synthesized Compounds

compd	<i>R_m</i> (0) ^a	hA ₁ <i>K_i</i> (nM) ^b	hA _{2A} <i>K_i</i> (nM) ^c	hA _{2B} <i>K_i</i> (nM) ^d	hA ₃ <i>K_i</i> (nM) ^e	hA ₁ /hA ₃	hA _{2A} /hA ₃	hA _{2B} /hA ₃
3	3.06 ± 0.06	250 ± 23	60 ± 10	200 ± 16	0.04 ± 0.009	6250	1500	5000
4	2.29 ± 0.05	350 ± 22	100 ± 12	250 ± 24	0.01 ± 0.005	35000	10000	25000

^a The *R_m* values of **3** and **4** were measured with a mobile phase of different concentrations of MeOH/H₂O. *R_m* values are reported as theoretical at 0% organic solvent in the mobile phase (*R_m*(0)). ^b Displacement of specific [³H]DPCPX²⁶ binding at human A₁ receptors expressed in CHO cells (*n* = 3–6). ^c Displacement of specific [³H]ZM 241385²⁶ binding at human A_{2A} receptors expressed in HEK-293 cells. ^d Displacement of specific [³H]DPCPX binding at human A_{2B} receptors expressed in HEK-293²⁶ cells (*n* = 3–6). ^e Displacement of specific [³H]MRE3008-F20²⁶ binding at human A₃ receptors expressed in HEK-293 cells. Data are expressed as *K_i* ± SEM (*n* = 3–6). The affinity of **4** at the rat adenosine receptor in RBL-2H3 cells was also determined using previously reported methods.²¹ *K_i* values (*n* = 3) were 226 ± 50 nM (A₁), 97.6 ± 26.2 nM (A_{2A}), and >1 μM (A₃).

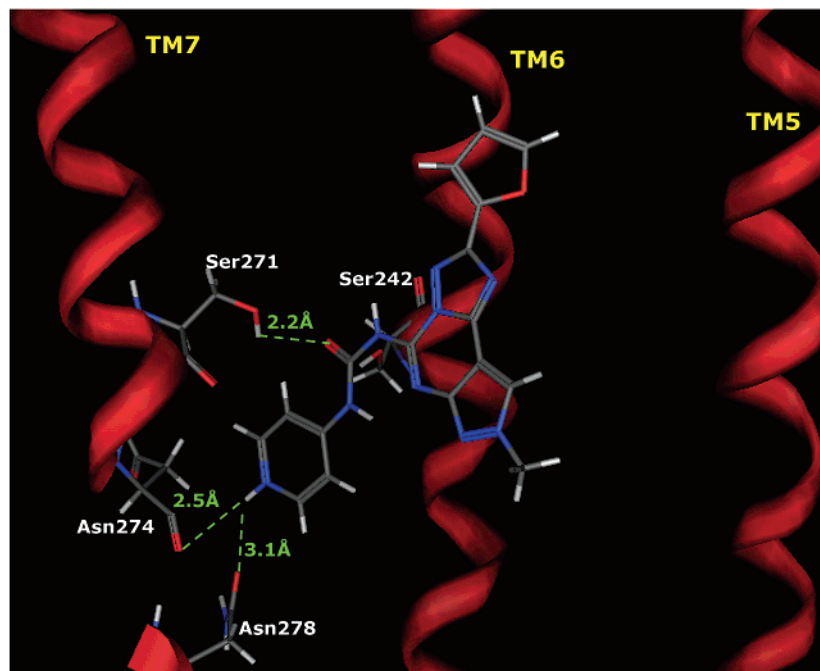


Figure 1. Side view of the human A_3 -**4** complex model. The side chains of the important residues in proximity to the docked **4** molecule are highlighted and labeled.

speculate that a difference in their relative dissociation/association rates during the biological assay could be a reasonable explanation. These results not only represent the first example of a highly potent, selective, and water-soluble human A_3 adenosine antagonist but strongly suggest an involvement of the pyridine nitrogen in the receptor recognition. Using a homology modeling approach based on the crystal structure of the rhodopsin²² as a template, we have built an improved model of the transmembrane helical domains (TMs) of the human A_3 receptor,²³ which can be considered a further refinement of the hypothetical binding site for A_3 receptor antagonists already proposed.^{13,22,24} As shown in Figure 1, after the Monte Carlo/annealing sampling, we propose that the hypothetical binding site of **4** is surrounded by TMs 3, 5, 6, and 7, with the furan ring pointing toward the extracellular environment. Similar to conformational results already described for other pyrazolotriazolopyrimidines,^{12,13,22} the lowest energy conformation of **4** featured the carbamoyl moiety in the 5-position surrounded by four polar amino acids: Ser242 (TM6), Ser271 (TM7), His274, and Ser275 (TM7).

Accordingly, this region seemed to be critical for the recognition of this class of antagonists. Moreover, additional strong electrostatic interactions appeared to occur between the positively charged pyridinium moiety of **4** and the carbonyl oxygen atoms of Asn274 ($N^+H \cdots O=C$ distance = 2.5 Å) and Asn278 ($N^+H \cdots O=C$ distance = 3.1 Å), both located on TM7. These electrostatic interactions might be responsible for the increase of the affinity in the protonated form, i.e., the hydrochloride derivative **4**. Interestingly, these two asparagine residues (Asn274 and Asn278) are largely conserved among a number of GPCRs. It should be noted, however, that this human A_3 receptor model, based on the TM region of the receptor, is not able to clearly explain the corresponding increase of selectivity for this subtype even taking into account which of the important TM residues are different in the other receptor subtypes.

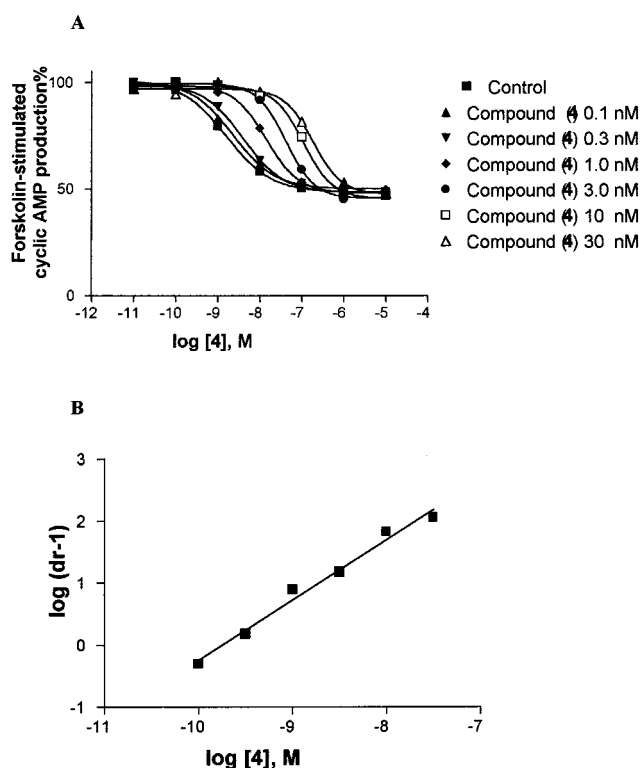


Figure 2. Effects of **4** on the inhibition of cyclic AMP production induced by the agonist CI-IB-MECA in human A_3 adenosine receptor-expressing CHO cells (A) and Schild analysis (B) of the data. The procedures used are described in Supporting Information. The data shown in panel A were derived from one experiment performed in duplicate and are typical of three independent experiments giving similar results. The K_B value for antagonism by **4** was calculated from three independent experiments.

However, as already reported by Jacobson and co-workers, multiple regions of the adenosine receptors, including a segment of the second extracellular loop, are involved in ligand recognition.²⁵ Other investigations

are in progress in our lab to better describe the role of the extracellular domain on the ligand recognition process. For confirmation of the high potency of this compound in a functional assay, the inhibition of cAMP²⁶ generation by Cl-IB-MECA in membranes of CHO cells stably transfected with the human A₃ receptor was evaluated.

Consistent with its binding affinity, **4** showed an IC₅₀ value of 0.7 ± 0.06 nM, compared to an IC₅₀ of 2.10 ± 0.21 nM for **1**. A Schild analysis of the antagonism of the effects of **4** on Cl-IB-MECA²⁶-induced inhibition of forskolin-stimulated cAMP was carried out (Figure 2).¹⁹ A K_B value of 0.20 ± 0.03 nM was calculated, thus demonstrating **4** to be the most potent antagonist of the human A₃ receptor ever reported.

Conclusions. The present study revealed a novel, potent, selective, and most importantly, water-soluble hA₃ adenosine receptor antagonist. This derivative featured a basic 4-pyridylcarbonyl moiety at the N5 position of the pyrazolotriazolopyrimidine nucleus, and the corresponding hydrochloride salt **4** displayed a K_i value of 0.01 nM at the hA₃ and selectivities versus the other adenosine receptor subtypes ranging from 10 000 to 35 000. This increase of affinity compared to neutral arylcarbamate derivatives could be attributed in receptor modeling to strong electrostatic interactions between the pyridinium moiety of **4** and the side chain carbonyl oxygen atoms of Asn274 and Asn278, both located on TM7. In view of the potency, selectivity, and water solubility, **4** could be an ideal candidate for pharmacological and clinical investigation of the hA₃ adenosine receptor subtype.

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Supporting Information Available: Experimental details. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- Fredholm, B. B.; Ijzerman, A. P.; Jacobson, K. A.; Klotz, K. N.; Linden, J. International Union of Pharmacology. XXV. Nomenclature and classification of adenosine receptors. *Pharmacol. Rev.* **2001**, *53*, 527–552.
- Ralevic, V.; Burnstock, G. Receptors for purines and pyrimidines. *Pharmacol. Rev.* **1998**, *50*, 413–492.
- Salvatore, C. A.; Jacobson, M. A.; Taylor, H. E.; Linden, J.; Johnson, R. G. Molecular cloning and characterization of the human A₃ adenosine receptor. *Proc. Natl. Acad. Sci. U.S.A.* **1993**, *90*, 10365–10369.
- Jacobson, K. A.; Suzuki, F. Recent developments in selective agonists and antagonists acting at purine and pyrimidine receptors. *Drug. Dev. Res.* **1996**, *39*, 289–300.
- Abbraccio, M. P.; Brambilla, R.; Kim, H. O.; von Lubitz, D. K. J. E.; Jacobson, K. A.; Cattabeni, F. G-protein-dependent activation of phospholipase-C by adenosine A₃ receptor in rat brain. *Mol. Pharmacol.* **1995**, *48*, 1038–1045.
- Ali, H.; Choi, O. H.; Fraundorfer, P. F.; Yamada, K.; Gonzaga, H. M. S.; Beaven, M. A. Sustained activation of phospholipase-D via adenosine A₃ receptors is associated with enhancement of antigen-ionophore-induced and Ca²⁺-ionophore-induced secretion in a rat mast-cell line. *J. Pharmacol. Exp. Ther.* **1996**, *276*, 837–845.
- Ramkumar, V.; Stiles, G. L.; Beaven, M. A.; Ali, H. The A₃ adenosine receptors is the unique adenosine receptor which facilitates release of allergic mediators in mast cells. *J. Biol. Chem.* **1993**, *268*, 16887–16890.
- Jacobson, K. A.; Moro, S.; Kim, Y. C.; Li, A. H. A₃ adenosine receptors: protective vs. damaging effects identified using novel agonists and antagonists. *Drug Dev. Res.* **1998**, *45*, 113–124.
- Brambilla, R.; Cattabeni, F.; Ceruti, S.; Barbieri, D.; Franceschi, C.; Kim, Y.; Jacobson, K. A.; Klotz, K. N.; Lohse, M. J.; Abbraccio, M. P. Activation of the A₃ adenosine receptor effects cell cycle progression and cell growth. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2000**, *361*, 225–234.
- Baraldi, P. G.; Cacciari, B.; Romagnoli, R.; Merighi, S.; Varani, K.; Borea, P. A.; Spalluto, G. A₃ adenosine receptor ligands: history and perspectives. *Med. Res. Rev.* **2000**, *20*, 103–128.
- Baraldi, P. G.; Cacciari, B.; Romagnoli, R.; Spalluto, G.; Klotz, K.-N.; Leung, E.; Varani, K.; Gessi, S.; Merighi, S.; Borea, P. A. Pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine derivatives as highly potent and selective human A₃ adenosine receptor antagonists. *J. Med. Chem.* **1999**, *42*, 4473–4478.
- Baraldi, P. G.; Cacciari, B.; Romagnoli, R.; Spalluto, G.; Moro, S.; Klotz, K. N.; Leung, E.; Varani, K.; Gessi, S.; Merighi, S.; Borea, P. A. Pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine derivatives as highly potent and selective human A₃ adenosine receptor antagonists: Influence of the chain at N⁸ pyrazole nitrogen. *J. Med. Chem.* **2000**, *43*, 4768–4780.
- Baraldi, P. G.; Cacciari, B.; Moro, S.; Spalluto, G.; Pastorin, G.; Da Ros, T.; Klotz, K.-N.; Varani, K.; Gessi, S.; Borea, P. A. Synthesis, Biological Activity, and Molecular Modeling Investigation of New Pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine Derivatives as Human A₃ Adenosine Receptor Antagonists. *J. Med. Chem.* **2002**, *45*, 770–780.
- Singha, C. N.; Dixit, N.; Sathyanarayana, D. N. ¹H and ¹³C NMR spectra of some unsymmetric *N,N*-dipyrilidyl ureas: spectral assignments and molecular conformations. *J. Chem. Soc., Perkin Trans. 2* **1997**, 157–162.
- Curtius, T.; Mohr, E. Transformation of nicotinic acid to beta-amidopyridine. *Ber.* **1898**, *31*, 2493–2495.
- Hyden, S.; Wilbert, G. Pyridine isocyanates. *Chem. Ind. (London)* **1967**, *33*, 1406–1407.
- Biagi, G. L.; Barbaro, A. M.; Sapone, A.; Borea, P. A.; Varani, K.; Recanatini, M. Study of lipophilic character of serotonergic ligands. *J. Chromatogr., A* **1996**, *723*, 135–143.
- Lohse, M. J.; Klotz, K.-N.; Lindernborn-Fotinos, J.; Reddington, M.; Schwabe, U.; Olsson, R. A. 8-Cyclopentyl 1,3-dipropylxanthine DPCPX a selective high affinity antagonist radioligand for A₁ adenosine receptors. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1987**, *336*, 204–210.
- Ongini, E.; Dionisotti, S.; Gessi, S.; Irenius, E.; Fredholm, B. B. Comparison of CGS 15943 and SCH 58261 as antagonist at human A₃ adenosine receptors. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1999**, *359*, 7–10.
- Varani, K.; Merighi, S.; Gessi, S.; Klotz, K. N.; Leung, E.; Baraldi, P. G.; Cacciari, B.; Spalluto, G.; Borea, P. A. [³H]MRE3008-F20: a novel antagonist radioligand for the pharmacological and biochemical characterization of human A₃ adenosine receptors. *Mol. Pharmacol.* **2000**, *57*, 968–975.
- (a) Jacobson, K. A.; Gallo-Rodriguez, C.; Melman, N.; Fischer, B.; Maillard, M.; van Bergen, A.; van Galen, P. J. M.; Karton, Y. Structure–activity relationships of 8-styrylxanthines as A₂-selective adenosine antagonists. *J. Med. Chem.* **1993**, *36*, 1333–1342. (b) Ji, X.-D.; Gallo-Rodriguez, C.; Jacobson, K. A. A selective affinity label for A₃ adenosine receptors. *Biochem. Biophys. Res. Commun.* **1994**, *203*, 570–576.
- Baraldi, P. G.; Cacciari, B.; Romagnoli, R.; Moro, S.; Ji, X.-D.; Jacobson, K. A.; Gessi, S.; Borea, P. A.; Spalluto, G. Fluorosulfonyl- and bis-(β-chloroethyl)amino-phenyl functionalized pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine derivatives as irreversible antagonists at the human A₃ adenosine receptor: molecular modeling studies. *J. Med. Chem.* **2001**, *44*, 2735–2742.
- Palczewski, K.; Kumasaka, T.; Hori, T.; Behnke, C. A.; Motoshima, H.; Fox, B. A.; Trong, I. L.; Teller, D. C.; Okada, T.; Stenkamp, R. E.; Yamamoto, M.; Miyano, M. Crystal structure of rhodopsin: a G protein-coupled receptor. *Science* **2000**, *289*, 739–745.
- Moro, S.; Li, A. H.; Jacobson, K. A. Molecular modeling studies of human A₃ adenosine antagonists: structural homology and receptor docking. *J. Chem. Inf. Comput. Sci.* **1998**, *38*, 1239–1248.
- Olah, M. E.; Jacobson, K. A.; Stiles, G. L. Role of the second extracellular loop of adenosine receptors in agonist and antagonist binding. Analysis of chimeric A₁/A₃ adenosine receptors. *J. Biol. Chem.* **1994**, *269*, 24692–24698.
- Abbreviations: DPCPX, 8-cyclopentyl-1,3-dipropylxanthine; [¹²⁵I]-AB-MECA, [¹²⁵I]-1-[6-[[[(4-amino-3-iodophenyl)methyl]amino]-9*H*-purin-9-yl]-1-deoxy-*N*-methyl-β-*D*-ribofuranuronamide; THF, tetrahydrofuran; CHO, Chinese hamster ovary; HEK, human embryonic kidney; MRE3008-F20, 5-[[[(4-methoxyphenyl)amino]carbonyl]amino-8-propyl-2-(2-furyl)-pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine; ZM 241385, 5-amino-7-(2-phenylethyl)-2-(2-furyl)pyrazolo[4,3-*e*]1,2,4-triazolo[1,5-*c*]pyrimidine; Cl-IB-MECA, 2-chloro-3-iodobenzyl-5'-(*N*-methylcarbamoyl)adenosine; cAMP, cyclic adenosine-5'-monophosphate.